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Brian Greenwood and Kevin De Cock

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6 Environmental factors, the immune system and the susceptibility to infection

Luc Kestens and Guido Vanham

Institute of Tropical Medicine, Antwerp, Belgium

Individual susceptibility to infection, disease and death is influenced by various factors such as host genotype, age, psychological state, virulence of the infectious agent and also by ecological, socio-economical and cultural changes. The interplay between these internal and external factors is complex and their relationship to new or resurgent infections is often very poorly understood. It is known that environmental factors can impair the immune system. To what extent these immunological changes modify susceptibility to infection is more difficult to assess and is often speculative.

In this paper, the diversity of the immune response to infectious agents and the impact of external factors such as malnutrition and environmental 'modulators' on these responses are reviewed. As an illustrative example, susceptibility to infection with the human immune deficiency virus (HIV) will be discussed, as well as the way in which this virus, as a recently introduced 'environmental' factor itself, has affected the reappearance of other infectious diseases. HIV changes host susceptibility to tuberculosis, toxoplasmosis and herpes simplex but not to streptococcal infections or malaria. To explain this paradox, it is essential to understand the mechanism of protective immunity to these infectious agents and the nature of the immune defects caused by HIV itself.

The immune response to infectious agents

When an infectious agent invades the body, the immune system activates innate and acquired (adaptive) immune responses (Figure 6.1). Innate immune responses are not antigen-specific, do not 'mature' upon repeated exposure, and require no 'education'. They constitute the first line of immune defence and involve humoral

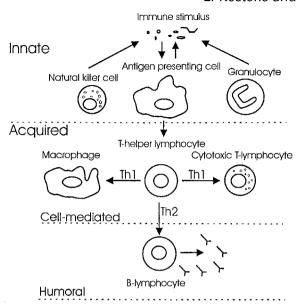


Figure 6.1 Immune responses to infectious agents comprise innate or non-specific responses and acquired or antigen-specific responses. The type of immune response is determined primarily by the nature of the infectious agent. Obligate intracellular viruses and parasites induce a cell-mediated immune response which is facilitated by cytokines which enhance cell-mediated immunity (Th-1 cytokines). Extracellular bacteria and parasites are neutralized mainly by a humoral immune response (antibodies, complement, etc.) which is driven by cytokines which enhance humoral immune responses (Th-2 cytokines)

components such as complement proteins, acute-phase proteins and other inflammatory mediators such as interferons and interleukins, as well as cellular components such as neutrophils, platelets, various tissue cells, mononuclear phagocytes and natural killer cells. On the other hand, adaptive immune responses are highly specific for a particular pathogen and improve with each re-exposure to the same pathogen. The adaptive immune response 'learns' and 'remembers' the infectious agent and can often prevent it from causing disease on a subsequent encounter. Cells which are central to adaptive immune responses are T-lymphocytes (T-cells) and B-lymphocytes (B-cells). B-cells produce antibodies whereas T-cells have another range of activities. Some subsets of T-cells are involved in helping B-cells to produce antibodies directed against extracellular pathogens (T-cell-dependent antibody production) whereas others interact with phagocytic and cytolytic cells and instruct them to destroy intracellular pathogens. Adaptive immune responses are generated in the context of the major histocompatibility complex (MHC) and are thus dependent on the genetic constitution of the host.

Mechanisms of protective immunity depend largely upon the nature of the infectious agents. Viruses are obligate intracellular organisms and require different immune responses than extracellular bacteria and parasites. As a rule, humoral immune responses are very efficient in neutralizing extracellular microbes whereas virus-infected cells and intracellular parasites can be destroyed only by cytolytic effector cells.

Host susceptibility or resistance to infection is determined, at least partially, by the type of immune response that the host develops. For instance, protective immune responses to intracellular viruses, bacteria and parasites such as cytomegalovirus, influenza virus, *Listeria monocytogenes*, *Mycobacterium leprae*, *M. tuberculosis*, *Leishmania* spp. and *Trypanosoma cruzi* are achieved predominantly through an adaptive cell-mediated immune response (Th1), whereas susceptibility to some of these agents is seen in individuals who mount a predominantly humoral immune (Th2) response.

Environmental factors can selectively interfere with either one or with both arms of the immune response and thus change the host susceptibility to infectious agents. Host factors which influence the immune response include genetic determinants and age. Environmental factors that can modulate the immune response include malnutrition, immunosuppressants, co-infections, mental stress, depression and exposure to pollutants and immunotoxins (Table 6.1).

Effect of the nutritional status on the immune response

Nutritional status is a well recognized determinant of immunocompetence. Nutritional disorders, affecting several hundred million people worldwide, can increase morbidity and mortality from many infections (Chandra and Newberne, 1977). Among the factors that determine nutritional status are food quality, food quantity and digestive efficiency. In addition, infection itself can deprive the body of nutrients and contribute to immunodeficiency, thus establishing a vicious circle (Storey, 1993). At one end of the spectrum of malnutrition are the gross changes seen in protein-energy malnutrition (PEM). More subtle changes are observed when specific nutrients like certain minerals and trace elements are selectively lacking.

Several components of the immune system are impaired in children who suffer from severe malnutrition. In PEM, the observed immunological defects are partially due to the effect of malnutrition on the lymphoid tissues, which are particularly susceptible as a result of their rapid rate of turnover and synthesis of immunomodulating proteins. Thymus, spleen, lymph nodes and Peyer's patches are altered in size and structure in children with severe PEM (Chandra, 1992).

Skin test responses to recall-antigens (delayed hypersensitivity skin responses), a useful *in vivo* measure of cell-mediated immunity, are reduced. Complete anergy (non-responsiveness) to a battery of different antigens is sometimes seen. A profound reduction of CD4+ T-helper cells is found in PEM, resulting in a

Table 6.1 Environmental factors which can affect the immune response

Environmental factor	Immunological impairment	Mechanism	Further reading
Malnutrition Protein energy malnutrition (PEM)	IFN-y and IL-2 production, phagocytic activity, Atrophy of lymphoid complement pathway, mucosal IgA response, organs DTH, T-LPR	Atrophy of lymphoid organs	Chandra, 1992; Chandra and Kumar, 1994 (overview)
Irace element deficiency Fe, Zn, Cu, Se	T-LPR (mitogens), NK, CTL, intracellular killing Reduced antioxidant of bacteria by phagocytes, lymphokine function (metalloenzy production, neutrophil function dismutase (Fe), superox dismutase (Zn/Cu), glutathione peroxidase (Se)	Reduced antioxidant function (metalloenzymes catalase (Fe), superoxide dismutase (Zn/Cu), glutathione peroxidase (Se))	Good and Lorenz, 1992; Bendich, 1993; Weiss <i>et al</i> , 1995; Harbige, 1996
Vitamin deficiency Vitamin A	CMI, T-cell-dependent antibody production,	Unknown	Rumore, 1993
Vitamin B6 and B12	DTH, T-LPR, CTL	Thymic atrophy, B6 and B12 mediate DNA	Miller, 1992 (overview)
Vitamin C, Vitamin E	DTH, T-LPR, neutrophil bactericidal function, mucosal surface integrity	synthesis Antioxidant, co-factor in hydroxylation of proline and lysine (collagen synthesis)	Harbige, 1996; Chew, 1995
Neurological stress Mental stress Severe depression	NK, T-LPR	(?) Neuropeptide receptors Cohen, 1995; Cohen and on immune cells Herbert, 1996; Glaser <i>et a</i> 1992; Irwin, 1988; Maes <i>et a</i> 1, 1991	Cohen, 1995; Cohen and Herbert, 1996; Glaser <i>et al</i> , 1992; Irwin, 1988; Maes <i>et al</i> , <i>al</i> , 1991

Immune suppressants			
Azathioprine	CMI and humoral immunity (blocks DNA synthesis)	Interferes with purine biosynthesis	Sigal and Dumont, 1993 (overview)
Steroids	CMI, IFN-y-antagonist, anti-inflammatory molecule	Steroid receptor on immune cells	Lew et al, 1988
Cyclosporin, FK-506	CMI (blocks lymphocyte activation)	Blocks IL-2 gene transcription	Sigal and Dumont, 1992
Rapamycin	CMI (blocks lymphocyte activation)	Blocks intracellular signalling through IL-2-R?	Sigal and Dumont, 1992
Environmental pollution Pesticides			
PCDDs, PCBs, PCDFs	NK, T-LPR, DTH, MLR, primary antigen-specific responses	TCDD binding to a de Swart et al, 1 cytosolic protein, the aryl Ross et al, 1995 hydrocarbon (Ah)	de Swart et al, 1994, 1995; Ross et al, 1995
		receptor, resemblance to TCDDs	
Ultraviolet B radiation (stratospheric ozone depletion)	DTH responses in the skin	Impairs function of antigen-presenting cells of 1996 the skin (Langerhans cells)	Morison, 1989; Patz <i>et al</i> , 1996
CMI = cell-mediated immini	CMI = cell-mediated immunity NK = natural killer cells: $CTL = cytotoxic$ T-lymphocytes: DTH = delayed type hypersensitivity: MTR = mixed	shocytes: DTH = delayed type hyr	persensitivity: MT.R = mixed

CMI = cell-mediated immunity; NK = natural killer cells; CTL = cytotoxic T-lymphocytes; DTH = delayed type hypersensitivity; MLK = mixed lymphocyte reaction; T-LPR = T-lymphocyte proliferative responses; PCDDs = polychlorinated dibenzo-p-dioxins; TCDDs = 2,3,7,8-tetrachloro-dibenzo-p-dioxin; PCBs = polychlorinated biphenyls; PCDFs = polychlorinated dibenzo-pracognans

significant reduction of T-helper (Th1) activity which explains, to a large extent, the alteration of cellular functions seen in children with PEM. Humoral (Th2) responses are less affected, as shown by near normal serum antibody responses after immunisation with common antigens. Nevertheless, mucosal IgA responses can be seriously depressed, enhancing the risk of mucosal infections (Chandra and Kumari, 1994), which, by resulting in diarrhoea, can further aggravate nutritional status and hence immune competence.

Some minerals, such as potassium, sodium, calcium, phosphorus and magnesium, are present in large amounts in the body. Others, such as selenium, zinc, iron and copper, are required only in small quantities, but nutritional deficiency of those trace elements can occur if available food lacks diversity. Enzyme and metabolic function may be significantly reduced by deficiencies in trace elements and vitamins. The most thoroughly studied trace element and vitamin deficiencies are those involving zinc, copper, iron, selenium, and vitamins A, B and E. Iron deficiency is the most common single nutrient deficiency, occurring in both developed and underdeveloped countries. It is characterized by reduced intracellular killing of bacteria by phagocytes, decreased T-cell numbers, reduced lymphocyte transformation to mitogens and by lowered lymphokine production (Weiss et al, 1995). The cellular basis of these various effects is reduced activity of ribonucleotidyl reductase (explaining the decreased proliferative responses depending on DNA and RNA synthesis) and decreased myeloperoxidase activity and hydroxyl radical production (explaining the deficient intracellular killing of bacteria by phagocytes). Zinc deficiency is associated with a depressed antibody production to heterologous antigens, reduced lymphocyte proliferative responses to mitogens, and depressed polymorphonuclear neutrophil, natural killer and cytotoxic T-cell function (Good and Lorenz, 1992). Selenium is necessary in sufficient amounts for protective immune responses to viruses. A critically low selenium level impairs T-cell functions and decreases natural killer cell activity (Harbige, 1996). Zinc, copper and iron are important in the antioxidant activity of some metallo-enzymes which neutralise intracellular free radicals. Free radicals and reactive oxygen species are produced by immune cells to destroy invading pathogens whereas the antioxidative system ensures that the formation of free radicals in living cells does not result in cellular damage (Bendich, 1993).

Deficiency of certain vitamins can also adversely affect immune functions. Vitamin A deficiency depresses cell-mediated immunity and T-cell-dependent antibody production, including secretory IgA production essential for the establishment of immunity to mucosal pathogens (Rumore, 1993). Vitamin E and C are naturally occurring antioxidant nutrients and enhance immunity, at least in part, by maintaining the functional and structural integrity of important immune cells (Chew, 1995; Harbige, 1996).

Immunosuppressants and susceptibility to infection

Immunosuppressants can increase the risk of infection significantly. Immunosuppressive agents can be grouped into several categories. Traditional immunosuppressives interfere with cellular metabolism and have a significant toxic effect. Examples of such antiproliferative agents are cyclophosphamide, methotrexate and azathioprine. They can cause non-specific suppression of both humoral and cell-mediated immune responses. Glucocorticosteroids suppress immune and inflammatory responses by inhibiting the expression of inflammatory mediators such as arachidonic acid metabolites and IL-1 (Lew et al, 1988). Another large group of agents inhibit signal transduction events at the cell surface or within the cell, resulting in non-specific immunosuppression and increased susceptibility to infection, although this is less drastic than that caused by the antiproliferative agents. Well known examples include cyclosporin A, FK-506 and rapamycin. They act as inhibitors of T-cell activation and interfere with the regulation of IL-2 gene transcription (CsA, FK506) or IL-2 receptor signal transduction (rapamycin) (Sigal and Dumont, 1992).

Mental stress, clinical depression and the immune system

Many studies have examined the relationship between mental stress, clinical depression and immunity (Cohen and Herbert, 1996). Emotional stress is often followed by increased susceptibility to bacterial and viral infections, at least for the less serious infectious diseases such as colds, influenza and herpes virus infection (Kiecolt-Glaser and Glaser, 1991; Cohen, 1995). The ability of mental stress to influence an immune response to a primary antigen has been well illustrated in a study of a group of students who were given a recombinant hepatitis B vaccine during a three-day academic examination period (Glaser et al, 1992). Those students who seroconverted after the first injection were significantly less stressed and anxious at the time of vaccination than those who did not seroconvert. Stress-associated reductions of important immunological anti-viral responses, such as those involving cytotoxic T-lymphocytes and natural killer cells, have also been observed in asymptomatic HIV-infected subjects (Evans et al, 1995). Similar observations were made in patients with severe depression (Irwin, 1988; Maes et al, 1991). Apparently, the immune system can be affected directly by innervation or by a neuro-endocrine cascade and vice versa. This bidirectional communication between the immune system and the central nervous system is achieved by means of common receptors and biologically active substances such as cytokines and neuropeptides (Savino and Dardenne, 1995).

Effect of environmental pollutants on the immune system

Because of the wide use of pesticides for domestic and industrial purposes, their potential immunotoxic effects are a matter of major concern for public health. Evidence that pesticides can severely impair immune functions in humans is scarce. Contact hypersensitivity is a well defined, although rare, consequence of exposure to pesticides, but immunologically mediated systemic reactions have been described only as debatable case reports (Vial et al, 1996). Studies in laboratory animals have shown that the mammalian immune system can be affected adversely by a variety of chemical agents (Vos and Luster, 1989; Saboori and Newcombe, 1992; Luster and Rosenthal, 1993; Lai et al, 1994) but, in most cases, these studies focused on acute immunotoxicity caused by relatively high levels of exposure. Potentially immunotoxic chemicals such as polychlorinated biphenyls (PCBs) and polychlorinated dibenzo-p-dioxins (PCDDs), dibenzofurans (PCDFs), hexachlorobenzene (HCB), dieldrin, \(\beta\)-hexachloro-cyclohexane (\(\beta\)-HCH) and dichlorodiphenyl trichloro-ethane (DDT) are present in abundance in the marine environment. Top predators are known to accumulate high levels of some of these xenobiotics. When morbillivirus infections led to massive mortalities among harbour seals in Europe in the early 1990s, it was speculated that this was caused by the adverse effect of environmental chemicals on the immune system of these animals. In a recent study carried out by de Swart et al (1995), statistically significant changes in cellular immune responses were detected in harbour seals that were fed on environmentally contaminated herring from the Baltic Sea. Whether induced immunological changes were the major determinant that promoted the virus infection and caused mass mortalities among seals is difficult to prove.

The mechanism of immune suppression induced by the most extensively studied group of immunotoxic chemicals, TCDD and related compounds including PCDDs, PCDFs and PCBs, is thought to be mediated by binding to a cytosolic protein, the aryl hydrocarbon receptor (Holsapple *et al*, 1991). The toxicity of these chemicals is largely dependent on their stereochemical resemblance to TCDD, the chemical with the highest affinity for this receptor. Through their interaction with this receptor, they can activate key protein kinases that are involved in the growth factor signal-transduction pathway (Matsumura, 1995).

Climatic factors can influence susceptibility to infectious diseases as well. Ultraviolet radiation is a possible factor involved in climate-related immune dysfunction. Ultraviolet B radiation induces selective biological alterations in the skin, including suppression of normal immune responses, probably through its effect on Langerhans cells, the most important antigen-presenting cells of the skin (Morison, 1989). The pathogenic consequences of UVB radiation can be observed in the exacerbation of infectious diseases and development of skin cancer (Vermeer and Hurks, 1994). UV-mediated immune suppression may become more important in the future as a consequence of an increased flux of ultraviolet radiation (Patz et al, 1996).

Susceptibility to HIV and the effect of HIV on the immune system

Cellular immune responses to HIV and disease progression

Human immune deficiency virus (HIV) is characterized by its dramatic effect on the immune system. It infects and kills CD4+ T-helper lymphocytes, which play a pivotal role in the generation of an immune response (Figure 6.1). Once an individual becomes infected with HIV, the host reacts with a vigorous immune response against the virus. HIV-specific cytotoxic T-cells are generated, killing infected cells, and neutralizing antibodies are produced which limit the spread of cell-free virus. Nevertheless, the virus escapes from these vigorous but apparently inadequate immune responses, resulting in a chronically overactivated immune system which finally collapses and leaves the host in a state of severe acquired immune deficiency (AIDS). Lymph nodes are the main reservoir for HIV, and even during the asymptomatic stage of the infection billions of virus particles are produced every day and millions of CD4+ T-cells are destroyed (Fauci et al, 1993; Ho et al, 1995; Wei et al, 1995).

CD8+ T-cells are thought to play an important role in the immune defence against HIV. Large numbers of activated cytotoxic CD8+ T-cells (CTL) are generated early after infection and may slow down disease progression (reviewed by Autran et al, 1996). Nevertheless, the persistence of virus replication indicates the inability of CTL to eradicate HIV. Moreover, during the asymptomatic stage of the infection, vigorous polyclonal CTL responses directed against HIV are associated with the generation of a large number of virus variants (Ho et al, 1995; Wei et al, 1995) which are no longer recognised by CTL (Phillips et al, 1991; Haas et al, 1996) and which ultimately result in disease progression.

CD8+ T-cells are also able to control HIV replication without killing infected cells. This antiviral activity appears to be mediated by soluble factors such as chemokines (Cocchi et al, 1995) and CD8+ T-cell antiviral factor (CAF) (reviewed by Levy et al, 1996). CD8+ T-cells from asymptomatic HIV-infected subjects produce high levels of these suppressor factors and it has been suggested that they are important in preventing progression to clinical disease (Levy et al, 1996).

CD8+ T-cells are activated by Th1-like cytokines (IL-2, IFN-γ, TNF-α) whereas Th2-like cytokines (IL-4, IL-5, IL-6, IL-10, IL-13) tend to suppress cell-mediated immune responses. Studies have suggested that HIV disease progression is associated with a profound shift from a 'protective' cell-mediated (Th1) to a 'non-protective' humoral (Th2) immune response (Clerici and Shearer, 1993) but this hypothesis has generated a great deal of controversy due to discordant findings from different laboratories and is not accepted universally.

Susceptibility or resistance to infection with HIV

During the past five years, it has become clear that (i) a small group of HIV-seropositive individuals who have been infected for at least 10 years have

not progressed to AIDS (so-called long-term non-progressors or LTNP) and (ii) there exists a small group of HIV-seronegative individuals who have been exposed to the virus many times and yet have not seroconverted or become infected.

The majority of LTNP have high levels of HIV-specific CTL in their peripheral blood and in their lymph nodes (Pantaleo *et al*, 1995) and their CD8+T-lymphocytes produce large quantities of HIV suppressor factor (Levy *et al*, 1996) responses that are thought to play an important role in preventing HIV disease progression in LTNP.

Interestingly, HIV-specific cellular immune responses have been detected in HIV-seronegative individuals who have been exposed to the virus many times without being infected, suggesting that innate and/or naturally acquired immune responses to HIV may be protective in rare individuals (reviewed by Shearer and Clerici, 1996). Peripheral blood mononuclear cells obtained from different individuals are not equally permissive to HIV, and CD4+ T-cells taken from apparently HIV-'resistant' subjects produce more chemokines (RANTES, MIP-1 and MIP-1β) than do CD4+ T-cells obtained from HIV-susceptible individuals (Paxton et al. 1996). Although it had already been shown that these chemokines could prevent infection of monocytes by monocytotropic HIV strains (Cocchi et al, 1995), the precise mechanism remained unknown until the discovery of the HIV co-receptors. The CD4 molecule, which is expressed at the cell surface of helper T-cells and monocytes, was identified as the primary cell receptor for HIV many years ago and, although HIV binds tightly to CD4, expression of CD4 was found to be insufficient to allow HIV entry into non-human cells (Maddon et al, 1986). The discovery that the chemokine receptor CCR5 acts as a co-receptor for HIV (Alkhatib et al, 1996) explains why and how certain chemokines can block HIV infection. They interfere with HIV infection by binding to and blocking receptor availability to HIV. Another member of the chemokine-receptor family, CXCR4 (fusin), had already been identified earlier as a co-receptor for lymphotropic HIV variants (Feng et al, 1996).

Shortly after the discovery of the two co-receptors for HIV, Samson et al (1996), Dean et al (1996) and Liu et al (1996) described a polymorphism in the gene coding for CCR5. The presence of a mutation in this gene, a 32 base pair deletion, confers resistance or partial resistance to HIV infection. The gene product results in a non-functional receptor that does not support membrane fusion of HIV with its target cell. Population studies indicate that the homozygous defect is found in 1–2% of Caucasians, and the heterozygous defect in 13–16% of this population (Samson et al, 1996; Huang et al, 1996). So far, no HIV-infected Caucasians, homozygous for the mutation, have been found. Whether heterozygotes are also less susceptible to HIV infection is still controversial (Samson et al, 1996; Huang et al, 1996). This protective allele appears to be absent in black populations from Western and Central Africa and from populations in Japan (Samson et al, 1996; Huang et al, 1996). Since apparent resistance to HIV has also been observed in

these populations (Fowke et al, 1996), factors other than defective co-receptors must also be involved.

Environmental factors can affect susceptibility to HIV infection

Successful replication of HIV in CD4+ T-lymphocytes is determined by the state of activation of HIV-infected cells. Only activated cells produce large amounts of infectious virus particles. Therefore, concurrent infections, which stimulate the immune system, may enhance HIV replication in infected cells and accelerate HIV disease progression. In addition, infections may activate the immune system of HIV-seronegative subjects and render them more susceptible to infection with HIV. As a consequence, one might expect to find a higher susceptibility to HIV in areas where many other infectious diseases are prevalent, such as most developing countries. The rapid spread of HIV in developing countries may, in part, be accounted for by this phenomenon. Infections, such as tuberculosis (TB), that chronically activate the immune system could be especially damaging (Vanham et al, 1996). Mycobacterium tuberculosis (MTB) can activate the expression of HIV in latently infected monocytic cell lines, and monocytes obtained from TB patients support HIV replication better than monocytes from control persons (Toossi et al, 1993; Lederman et al, 1994). MTB increases HIV replication in peripheral blood mononuclear cells and this is correlated with the level of cellular activation which is a predominant characteristic of HIV-infected subjects (Kestens et al, 1992; 1994; Goletti et al, 1996). HIV replication was found to increase 5- to 160-fold during the acute phase of MTB disease (Goletti et al, 1996). These observations provide the underlying explanation for the clinical finding that infection with MTB can accelerate the clinical course of HIV (Whalen et al, 1995).

Both ulcerative and non-ulcerative sexually transmitted diseases (STDs) are known to increase susceptibility to HIV (Laga et al, 1993; Torian et al, 1995). Ulcerative STDs probably facilitate HIV entry through mucosal lesions, rather than by their effect on the immune system. The precise role of non-ulcerative STDs, such as gonorrhoeal, chlamydial and trichomonal infections, needs further elucidation.

Infections such as those caused by Schistosoma mansoni infection may drive the immune response towards a Th2-like profile and increase susceptibility to HIV infection. In mice infected with S. mansoni, suppression of Th1 reactivity by a dominant Th2 response has been shown to result in failure of virus-specific CD8+ T-cell responses to vaccinia virus (Actor et al, 1993). This suggests that helminth infections can influence immune responses to concurrent viral infections (Kullberg et al, 1992). Bentwich et al (1996) have shown a very high prevalence of helminthic and other infections associated with extreme immune dysregulation as well as a high prevalence of HIV-1 in Ethiopian immigrants to Israel. Although one of the striking characteristics of the AIDS epidemic in Africa is the way the

disease differs from the pattern seen in other areas, the reasons for this are not clear. It has been suggested that changes in the host immune response caused by endemic infections and mostly helminth infections could account for at least a part of this pattern (Bentwich *et al*, 1995).

Immunisation of HIV-seropositive patients with recall antigens temporarily enhances HIV replication in infected subjects (Stanley et al, 1996), raising the question of the advisability of vaccinations in HIV-infected subjects. However, the protection afforded in most cases by vaccination outweighs the potential risks from a transient increase in immune activation.

HIV changes the susceptibility to other infectious diseases

HIV induces progressive and selective immune defects in infected humans. Cell-mediated immune responses of the Th1 type which are required to confer protective immunity against pathogens are particularly compromised. Epidemiological data have demonstrated that HIV-infected individuals are more susceptible to MTB and that the HIV epidemic has a central role in the worldwide resurgence of this infection (Barnes et al, 1991). A study conducted by Burwen et al (1995) estimated that HIV-induced immunosuppression accounts for a minimum of 30% of the excess TB cases during the period 1985–1990 in the US. In Côte d'Ivoire, the incidence of tuberculosis was 1104 per 100 000 among HIV-infected persons in 1991 but only 96 per 100 000 in HIV-seronegative persons, a figure which is lower than the overall incidence measured in 1981 (155 per 100 000) (Richards et al, 1995). HIV affects not only the incidence of MTB but also its clinical presentation. Reactivation of pulmonary TB may occur early in HIV infection whereas extrapulmonary or atypical disease is seen in patients with profound HIV-induced immunodeficiency (Lucas and Nelson, 1994).

Visceral leishmaniasis (*Leishmania donovani*) is another opportunistic infection seen with increased frequency in patients infected with HIV who live in areas endemic for Leishmania. As a result of HIV, visceral leishmaniasis is becoming more important in non-endemic areas as well (Albrecht *et al*, 1996).

Toxoplasmosis, Pneumocystis carinii and Herpes simplex are classical examples of latent infections which can be reactivated by HIV. In contrast, leprosy (M. leprae), malaria (Plasmodium falciparum) and amoebiasis (Entamoeba histolytica), which are at least partially controlled by cell-mediated immunity and which should theoretically be more frequent in HIV-positive people than in HIV-negatives, do not appear to be more frequent or more aggressive in HIV-infected subjects. The reasons for this are unclear.

Conclusion

It is clear that a variety of environmental factors can disturb the integrity of the immune system in a direct or indirect manner. They can change the susceptibility

of the host to infection and they may have a role in the spread of new or resurgent infectious diseases. Numerous publications have demonstrated that many of these factors induce measurable changes in cellular and humoral immunity. The relative increase in susceptibility to infection depends on the nature of the infectious agent and on the type and degree of immune perturbation.

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References

- Actor JK, Shirai M, Kullberg MC, Buller RM, Sher A, Berzofsky JA. Helminth infection results in decreased virus-specific CD8+ cytotoxic T-cell and Th1-cytokine responses as well as delayed virus clearance. *Proceedings of the National Academy of Sciences of the USA*, 1993; **90**: 948–952
- Akhatib G, Combadiere C, Broder CC et al. CC CKRS: a RANTES, MIP-1 alpha, MIP-1 beta receptor as a fusion cofactor for macrophage-tropic HIV-1. Science, 1996; 272: 1955–1958
- Albrecht H, Sobottka I, Emminger C et al. Visceral leishmaniasis emerging as an important opportunistic infection in HIV-infected persons living in areas nonendemic for Leishmania donovani. Archives of Pathology and Laboratory Medicine, 1996; 120: 189–198
- Autran B, Hadida F, Haas G. Evolution and plasticity of CTL responses against HIV. Current Opinions in Immunology, 1996; 8: 546-553
- Barnes PF, Bloch AB, Davidson PT, Snider DE. Tuberculosis in patients with human immunodeficiency virus infection. New England Journal of Medicine, 1991; 324: 1644–1650
- Bendich A. Physiological role of antioxidants in the immune system. *Journal of Dairy Sciences*, 1993; **76**: 2789–2794
- Bentwich Z, Kalinkovich A, Weisman Z. Immune activation is a dominant factor in the pathogenesis of African AIDS. *Immunology Today*, 1995; **16**: 187–191
- Bentwich Z, Weisman Z, Moroz C, Bar-Yehuda S, Kalinkovich A. Immune dysregulation in Ethiopian immigrants in Israel: Relevance to helminth infections? *Clinical and Experimental Immunology*, 1996; **103**: 239–243
- Burwen DR, Bloch AB, Griffin LD, Ciesielski CA, Stern HA, Onorato IM. National trends in the concurrence of tuberculosis and acquired immunodeficiency syndrome. *Archives of Internal Medicine*, 1995; **155**: 1281–1286
- Chandra RK. Nutrition and immunoregulation. Significance for host resistance to tumors and infectious diseases in humans and rodents. *Journal of Nutrition*, 1992; **122**: 754–757
- Chandra RK, Kumari S. Nutrition and immunity: an overview. *Journal of Nutrition*, 1994; **124 (8 Suppl.)**: 1433S–1435S
- Chandra RK, Newberne PM (eds). Nutrition, Immunity and Infection: Mechanisms of Interactions. New York: Plenum Press, 1977
- Chew BP. Antioxidant vitamins affect food animal immunity and health. *Journal of Nutrition*, 1995; **125 (6 Suppl.)**: 1804S–1808S
- Clerici M, Shearer GM. A Th1-Th2 switch is a critical step in the etiology of HIV infection. Immunology Today, 1993; 14: 107–111
- Cocchi F, Devico AL, Garzino-Demo A, Arya SK, Gallo RC, Lusso P. Identification of RANTES, MIP-1 alpha, and MIP-1 beta as the major HIV-suppressive factors

- produced by CD8(+) T cells. Science, 1995; 270: 1811-1815
- Cohen S. Psychological stress and susceptibility to upper respiratory infections. American Journal of Respiratory and Critical Care Medicine, 1995; 152: S53–S58
- Cohen S, Herbert TB. Health psychology: psychological factors and physical disease from the perspective of human psychoneuroimmunology. *Annual Review of Psychology*, 1996; 47: 113–142
- de Swart RL, Ross PS, Vedder LJ et al. Impairment of immune function in harbor seals (*Phoca vitulina*) feeding on fish from polluted waters. Ambio, 1994; 23: 155-159
- de Swart RL, Ross PS, Timmerman HH et al. Impaired cellular immune response in harbour seals (*Phoca vitulina*) feeding on environmentally contaminated herring. Clinical and Experimental Immunology, 1995; 101: 480-486
- Dean M, Carrington M, Winkler C et al. Genetic restriction of HIV-1 infection and progression to AIDS by a deletion allele of the CKR5 structural gene. Hemophilia Growth and Development Study, Multicenter AIDS Cohort Study, Multicenter Hemophilia Cohort Study, San Francisco City Cohort ALIVE Study. Science, 1996; 273: 1856–1862
- Evans DL, Leserman J, Perkins DO et al. Stress-associated reductions of cytotoxic T lymphocytes and natural killer cells in asymptomatic HIV infection. American Journal of Psychiatry, 1995; 152: 543-550
- Fauci AS, Pantaleo G, Embretson J, Haase AT. Viral burden and HIV disease—reply. *Nature*. 1993; **364**: 291–292
- Feng Y, Broder CC, Kennedy PE, Berger EA. HIV-1 entry cofactor: functional cDNA cloning of a seven-transmembrane, G protein-coupled receptor. Science, 1996; 272: 872–877
- Fowke KR, Nagelkerke NJD, Kimani J et al. Resistance to HIV-1 infection among persistently seronegative prostitutes in Nairobi, Kenya. Lancet, 1996; 348: 1347–1351
- Glaser R, Kiecolt-Glaser JK, Bonneau RH, Malarkey W, Kennedy S, Hughes J. Stress-induced modulation of the immune response to recombinant hepatitis-B vaccine. *Psychosomatic Medicine*, 1992; **54**: 22–29
- Goletti D, Weissman D, Jackson RW et al. Effect of Mycobacterium tuberculosis on HIV replication. Role of immune activation. Journal of Immunology, 1996; 157: 1271-1278
- Good RA, Lorenz E. Nutrition and cellular immunity. International Journal of Immunopharmacology, 1992; 14: 361–366
- Haas G, Plikat U, Debre P et al. Dynamics of viral variants in HIV-I Nef and specific cytotoxic T lymphocytes in vivo. Journal of Immunology, 1996; 157: 4212-4221
- Harbige LS. Nutrition and immunity with emphasis on infection and autoimmune disease. *Nutrition and Health*, 1996; **10**: 285–312
- Ho DD, Neumann AU, Perelson AS, Chen W, Leonard JM, Markowitz M. Rapid turnover of plasma virions and CD4 lymphocytes in HIV-1 infection. *Nature*, 1995; 373: 123-126
- Holsapple MP, Morris DL, Wood SC, Snyder NK. 2,3,7,8-tetrachlorodibenzo-p-dioxininduced changes in immunocompetence: possible mechanisms. *Annual Review of Pharmacology and Toxicology*, 1991; 31: 73–100
- Huang Y, Paxton WA, Wolinsky SM et al. The role of a mutant CCR5 allele in HIV-1 transmission and disease progression. Nature Medicine, 1996; 2: 1240-1243
- Irwin M. Depression and immune function. Stress Medicine, 1988; 14: 95-103
- Kestens L, Vanham G, Gigase P et al. Expression of activation antigens, HLA-DR and CD38 on CD8 lymphocytes during HIV-1 infection. AIDS, 1992; 6: 793-797
- Kestens L, Vanham G, Vereecken C et al. Selective increase of activation antigens HLA-DR and CD38 on CD4(+)CD45RO(+) T lymphocytes during HIV-1 infection. Clinical and Experimental Immunology, 1994; 95: 436-441
- Kiecolt-Glaser JK, Glaser R. Stress and immune function in humans. In: Ader R, Felten

- DL, Cohen N (eds), *Psychoneuroimmunology*. San Diego, CA: Academic Press, 1991, pp 849–867
- Kullberg MC, Pearce EJ, Hieny SE, Sher A, Berzofsky JA. Infection with *Schistosoma mansoni* alters Th1-Th2 cytokine responses to a non-parasite antigen. *Journal of Immunology*, 1992; **148**: 3264–3270
- Laga M, Manoka A, Kivuvu M et al. Non-ulcerative sexually transmitted diseases as risk factors for HIV-1 transmission in women: results from a cohort study. AIDS, 1993; 7: 95-102
- Lai ZW, Kremer J, Gleichmann E, Esser C. 3,3',4,4'-tetrachlorobiphenyl (TCB) inhibits proliferation of immature thymocytes in fetal thymus organ culture. *Scandinavian Journal of Immunology*, 1994; **39**: 480–488
- Lederman MM, Georges DL, Kusner DJ, Mudido P, Giam CZ, Toossi Z. Mycobacterium tuberculosis and its purified protein derivative activate expression of the human immunodeficiency virus. Journal of Acquired Immune Deficiency Syndromes, 1994; 7: 727-733
- Levy JA, Mackewicz CE, Barker E. Controlling HIV pathogenesis: the role of the noncytotoxic anti-HIV response of CD8(+)T cells. *Immunology Today*, 1996; 17: 217–224
- Lew W, Oppenheim JJ, Matsushima K. Analysis of the suppression of IL-1 alpha and IL-1 beta production in human peripheral blood mononuclear adherent cells by a glucocorticoid hormone. *Journal of Immunology*, 1988; **140**: 1895–1902
- Liu R, Paxton WA, Choe S et al. Homozygous defect in HIV-1 coreceptor accounts for resistance of some multiply-exposed individuals to HIV-1 infection. Cell, 1996; 86: 367–377
- Lucas S, Nelson AM. Pathogenesis of tuberculosis in human immunodeficiency virus-infected people. In: Bloom BR (ed), *Tuberculosis: Pathogenesis, Protection and Control.* Washington, DC: American Society for Microbiology Press, 1994, pp 503–513
- Luster MI, Rosenthal GJ. Chemical agents and the immune response. *Environmental Health Perspectives*, 1993; **100**: 219–236
- Maddon PJ, Dalgleish AG, McDougal JS, Clapham PR, Weiss R, Axel R. The T4 gene encodes the AIDS virus receptor and is expressed in the immune system and the brain. *Cell*, 1986; 47: 333–348
- Maes M, Bosmans E, Suy E, Minner B, Raus J. A further exploration of the relationships between immune parameters and the HPA-axis activity in depressed patients. *Psychological Medicine*, 1991; 21: 313–320
- Matsumura F. Mechanism of action of dioxin-type chemicals, pesticides, and other xenobiotics affecting nutritional indexes. *American Journal of Clinical Nutrition*, 1995; 61: 695S-701S
- Miller LT. Vitamin B group and the immune system. In: Roit IM, Delves PJ (eds), Encyclopedia of Immunology. London: Academic Press, 1992, pp 1564–1565
- Morison W. Effects of ultraviolet radiation on the immune system in humans. *Photochemistry* and *Photobiology*, 1989; **50**: 515–524
- Pantaleo G, Menzo S, Vaccarezza M et al. Studies in subjects with long term nonprogressive human immunodeficiency virus infection. New England Journal of Medicine, 1995; 332: 209–216
- Patz JA, Epstein PR, Burke TA, Balbus JM. Global climate change and emerging infectious diseases. Journal of the American Medical Association, 1996; 275: 217-223
- Paxton WA, Martin SR, Tse D et al. Relative resistance to HIV-1 infection of CD4 lymphocytes from persons who remain uninfected despite multiple high-risk sexual exposures. Nature Medicine, 1996; 2: 412-417
- Phillips RE, Rowland-Jones S, Nixon DF et al. Human immunodeficiency virus genetic variation that can escape cytotoxic T-cell recognition. *Nature*, 1991; **354**: 453–459
- Richards SB, St Louis ME, Nieburg P et al. Impact of the HIV epidemic on trends in

- tuberculosis in Abidjan, Côte d'Ivoire. Tubercle and Lung Disease, 1995; 76: 11-16
- Ross PS, de Swart RL, Reijnders PJ, Van Loveren H, Vos JG, Osterhaus AD. Contaminant-related suppression of delayed-type hypersensitivity and antibody responses in harbor seals fed herring from the Baltic Sea. *Environmental Health Perspectives*, 1995; 103: 162–167
- Rumore MM. Vitamin A as an immunomodulating agent. Clinical Pharmacology and Therapeutics, 1993; 12: 506-514
- Saboori AM, Newcombe DS. Environmental chemicals with immunotoxic properties. In: Newcombe DS, Rose NR, Bloom JC (eds), Clinical Immunotoxicology. New York: Raven Press, 1992, pp 365-400
- Samson M, Libert F, Doranz BJ et al. Resistance to HIV-1 infection in Caucasian individuals bearing mutant alleles of the CCR-5 chemokine receptor gene. Nature, 1996; 382: 722-725
- Savino W, Dardenne M. Immune-neuroendocrine interactions. *Immunology Today*, 1995; **16**: 318-322
- Shearer GM, Clerici M. Protective immunity against HIV infection: has nature done the experiment for us? *Immunology Today*, 1996; 17: 21-24
- Sigal NH, Dumont FJ. Cyclosporin A, FK-506 and rapamycin: pharmacologic probes of lymphocyte signal transduction. *Annual Review of Immunology*, 1992; **10**: 519–560
- Sigal NH, Dumont FJ. Immunosuppression. In: Paul WE (ed), Fundamental Immunology. New York: Raven Press, 1993, pp 903-915
- Stanley SK, Ostrowski MA, Justement JS et al. Effect of immunization with a common recall antigen on viral expression in patients infected with human immunodeficiency virus type 1. New England Journal of Medicine, 1996; 334: 1222–1230
- Storey DM. Filariasis: nutritional interactions in human and animal hosts. *Parasitology*, 1993; 107: S147–S158
- Toossi Z, Sierra-Madero JG, Blinkhorn RA, Mettler MA, Rich EA. Enhanced susceptibility of blood monocytes from patients with pulmonary tuberculosis to productive infection with human immunodeficiency virus type 1. *Journal of Experimental Medicine*, 1993; 177: 1511–1516
- Torian LV, Weisfuse IB, Makki HA, Benson DA, DiCamillo LM, Toribio FE. Increasing HIV-1 seroprevalence associated with genital ulcer disease, New York City, 1990–1992. *AIDS*, 1995; **9**: 177–181
- Vanham G, Edmonds K, Qing L et al. Generalized immune activation in pulmonary tuberculosis: Co-activation with HIV infection. Clinical and Experimental Immunology, 1996; 103: 30–34
- Vermeer BJ, Hurks M. The clinical relevance of immunosuppression by UV irradiation. Journal of Photochemistry and Photobiology-B, Biology, 1994; 24: 149-154
- Vial T, Nicolas B, Descotes J. Clinical immunotoxicity of pesticides. *Journal of Toxicology* and Environmental Health, 1996; **48**: 215–229
- Vos JG, Luster MI. Immune alterations. In: Kimbrough RD, Jensen S (eds), Halogenated Biphenyls, Terphenyls, Naphthalenes, Dibenzodioxins and Related Products. Amsterdam: Elsevier Science Publishers BV, 1989, pp 295–322
- Wei X, Ghosh SK, Taylor ME et al. Viral dynamics in human immunodeficiency virus type 1 infection. Nature, 1995; 373: 117-122
- Weiss G, Wachter H, Fuchs D. Linkage of cell-mediated immunity to iron metabolism. *Immunology Today*, 1995; **16**: 495–500
- Whalen C, Horsburgh CR, Hom D, Lahart C, Simberkoff M, Ellner J. Accelerated course of human immunodeficiency virus infection after tuberculosis. *American Journal of Respiratory and Critical Care Medicine*, 1995; 151: 129–135