

Comparison of the direct faecal smear and two thick smear techniques for the diagnosis of intestinal parasitic infections

Dirk Engels^{1,2*}, Samuel Nahimana² and Bruno Gryseels³ ¹Belgian Technical Co-operation, Bujumbura, Burundi; ²Schistosomiasis Control Programme, Bujumbura, Burundi; ³Prince Léopold Institute of Tropical Medicine, Antwerp, Belgium

Abstract

Among 547 health centre patients in Burundi, the diagnostic performance of a glass coverslip modification of the Kato thick smear technique was compared with the combination of direct slide examination and the quick Kato-Katz method, currently recommended in basic health services, for the diagnosis of intestinal parasitic infections. The classical Kato-Katz method performed best for the diagnosis of common helminth infections, especially in combination with direct examination. For the diagnosis of protozoa, both trophozoites and cysts, the direct slide examination was superior to the glass coverslip technique. Despite its being a single and easy procedure, the glass coverslip technique could not be recommended as the method of choice for the diagnosis of intestinal parasites in basic health services.

Keywords: intestinal parasites, diagnosis, Kato-Katz method, faecal examination

Introduction

The Kato-Katz method (KATO & MIURA, 1954; KATZ *et al.*, 1972) is widely used for the field diagnosis of intestinal helminth infections. With this technique, a measured amount of stool (25-50 mg) is examined, approximately 10-20 times more than the quantity examined with the direct smear technique. It therefore considerably increases the diagnostic sensitivity and also permits a quantitative diagnosis. The Kato-Katz method is,

perform it. TEESDALE & AMIN (1976a, 1976b) and TEESDALE *et al.* (1985) have proposed a glass coverslip modification of the Kato thick smear method which can be examined immediately and is able to detect a wider range of intestinal parasites. We have tested, in 2 health centres in Burundi, the diagnostic performance of this method in comparison with the direct slide examination, the quick Kato-Katz method, and a combination of both techniques.

Table 1. Prevalences of common intestinal parasites detected by different techniques of faecal examination in 529 health centre patients in Burundi

Parasite	Coverslip technique	Prevalence (%)		
		Direct examination	Kato-Katz technique	Direct examination + Kato-Katz technique
Helminths, eggs/larvae				
<i>Schistosoma mansoni</i>	35.3	17.2***	38.6	40.8**
Hookworm	23.3	16.1***	48.2***	49.1***
<i>Ascaris lumbricoides</i>	3.6	3.0	4.2	4.2
<i>Strongyloides stercoralis</i>	2.8	1.7	0.4	1.7
<i>Trichuris trichiura</i>	1.3	0.9	4.9***	4.9***
<i>Taenia</i> spp.	0.4	0.6	0.4	0.8
<i>Enterobius vermicularis</i>	0.2	-	-	-
Protozoa, trophozoites				
<i>Enteromonas hominis</i>	22.9	36.1***	-	36.1***
<i>Giardia duodenalis</i>	18.5	25.0***	-	25.0***
<i>Entamoeba</i> spp.	16.8	29.1***	-	29.1***
<i>Trichomonas hominis</i>	5.5	8.3*	-	8.3*
<i>Chilomastix mesnili</i>	2.5	3.8	-	3.8
<i>Balantidium coli</i>	0.2	0.2	-	0.2
Protozoa, cysts				
<i>Entamoeba histolytica</i>	25.1	31.0**	-	31.0**
<i>Entamoeba coli</i>	5.5	8.1*	-	8.1*
<i>Giardia duodenalis</i>	1.1	2.1	-	2.1

*Significance of differences between the results indicated and the coverslip technique is shown thus: * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$; all other values were not statistically significantly different.

however, not suitable for the detection of hookworm, protozoa and filariform larvae (e.g., *Strongyloides stercoralis*). It also has the disadvantage of requiring a certain clearing time. ENGELS *et al.* (1993) have shown that, in combination with direct slide examination, a quick variant of this method (PETERS *et al.*, 1980) can considerably improve the diagnosis of *Schistosoma mansoni* and common intestinal helminths in basic health services. This variant method takes only 45 min and is also suitable for hookworm detection. But even this quick Kato-Katz method remains relatively tedious in a health centre setting and some centres are reluctant to

Materials and Methods

We asked 547 patients visiting 2 rural health centres to supply a stool specimen, regardless of their complaints. Each specimen was examined by means of the 3 techniques by the same microscopist, who was unaware of the origin of each slide. Three experienced microscopists participated in the study.

The direct examination (one slide per person) was carried out without staining. The 25 mg Kato-Katz slide (KATZ *et al.*, 1972; PETERS *et al.*, 1980) was also prepared without staining and examined after 45 min. The glass coverslip technique involved sieving the stool and placing a measured aliquot (25 mg) on a microscope slide (2.5 × 7.5 cm), using the same template as in the Kato-Katz method. One drop of normal saline was

*Present address, for correspondence: Avenue de Citeaux 13, 1348 Louvain-La-Neuve, Belgium.

Table 2. Comparison of the number of cases diagnosed by the combination of direct examination + Kato-Katz method, the coverslip technique, and both methods in 529 health centre patients in Burundi

Parasite	No. of cases diagnosed ^a			Total no. of cases
	DE+KK only	CT only	CT and DE+KK	
Helminths, eggs/larvae				
<i>Schistosoma mansoni</i>	64	35	152	251
Hookworm	147	10	113	270
<i>Ascaris lumbricoides</i>	4	1	18	23
<i>Strongyloides stercoralis</i>	4	10	5	19
<i>Trichuris trichiura</i>	22	3	4	29
<i>Taenia</i> spp.	3	1	1	5
<i>Enterobius vermicularis</i>	0	1	0	1
Protozoa, trophozoites				
<i>Enteromonas hominis</i>	96	26	95	217
<i>Giardia intestinalis</i>	56	22	78	156
<i>Entamoeba</i> spp.	88	22	69	179
<i>Trichomonas hominis</i>	25	11	19	55
<i>Chilomastix mesnili</i>	11	4	9	24
<i>Balantidium coli</i>	0	0	1	1
Protozoa, cysts				
<i>Entamoeba histolytica</i>	68	37	96	201
<i>Entamoeba coli</i>	26	12	17	55
<i>Giardia intestinalis</i>	5	0	6	11

^aCT = coverslip technique, DE = direct examination, KK = Kato-Katz method.

added to this aliquot before covering it with a glass coverslip (2.0×2.5 cm). The slide was then turned upside down on a piece of blotting paper and gentle but firm pressure was applied to spread the stool in a thin layer. The sample was examined immediately under the microscope at a magnification of ×100, or ×400 when more precise identification of parasites was required. As the aim of the study was to test the value of the different diagnostic methods in a health centre setting, only qualitative diagnosis was considered.

Statistical testing of the difference between prevalences observed by the different techniques was done by means of the McNemar test for paired proportions. When fewer than 10 cases had different outcomes (positive or negative) with the 2 techniques, the binomial distribution was used to calculate the exact value of *P*.

Results

All 3 methods of faecal examination were conducted on 529 people; the results are summarized in Table 1.

Fewer helminth infections were detected with the direct examination than with the glass coverslip technique. This difference was most marked for *S. mansoni* and hookworm. The Kato-Katz technique, especially when combined with direct slide examination, gave better results than the glass coverslip technique for all helminth infections, apart from strongyloidiasis and enterobiasis. Direct stool examination was more sensitive for the diagnosis of protozoa than the glass coverslip technique. This was true for both vegetative forms and cysts, and the difference was most marked for enteromoniasis, giardiasis and amoebiasis.

The prevalences of *S. mansoni*, hookworm and *Taenia* species determined by the combination of direct examination and the Kato-Katz method were higher than those determined by a Kato-Katz slide only. Twelve cases of schistosomiasis, 5 of hookworm infection, and 2 of taeniasis were diagnosed by the direct slide method but not by the Kato-Katz technique. The numbers of cases diagnosed by the glass coverslip technique, the combination direct slide Kato-Katz slide, and both methods are compared in Table 2.

Discussion

Compared with direct examination, the Kato-Katz

method is known to be more sensitive for the diagnosis of *S. mansoni*, *Ascaris*, *Trichuris* and *Taenia* infections. The quick variant of this method is also more sensitive for diagnosis of hookworm infection. Our results confirmed this, and also showed that a combination of both methods could further increase the sensitivity for infections with *S. mansoni*, hookworm and *Taenia* species.

The results also showed that the diagnostic performance of the combination of direct examination and a Kato-Katz slide was superior to the glass coverslip technique. This is particularly true for *S. mansoni* and hookworm, 2 important health problems in the area where the study was conducted. The glass coverslip technique performed better for strongyloidiasis only, a less common health problem in Burundi, and for enterobiasis, for which stool examination is not the diagnostic method of choice. Somewhat unexpected was the superior sensitivity of direct examination over the glass coverslip technique for the diagnosis of trophozoites and cysts of protozoa. This was possibly due to the fact that, without a clearing stage, the examination of a thick faecal smear is not easy. In fact, the microscopists complained about the difficulty of examining the glass coverslip preparations. The use of 2 microscope slides instead of a slide and a coverslip, as described by TEESDALE *et al.* (1985), may allow the stool to be spread out a little more, but it was impossible to examine such a 'glass sandwich' at magnification of ×400 with the microscopes routinely available in basic health services in Burundi.

The infections detected with different techniques were not necessarily the same (Table 2). This means that even the sensitivity of these 'improved' techniques is far from ideal, and health personnel should be aware that a person cannot with certainty be classified as uninfected on the basis of a single negative examination by one of these techniques.

The present study was intended to determine whether, in Burundi, the glass coverslip technique could replace, with advantage, the combination of direct and Kato-Katz examinations as the method of choice for the coprological diagnosis of intestinal parasites in basic health services. The glass coverslip technique has the advantage of being a single procedure, and would therefore considerably simplify the process and encourage its

adoption by health centres. Using the current procedure, the proportion of incomplete examinations is considerable in some centres, particularly in private health services and urban areas. This phenomenon seems to be related to both organizational aspects of health care and the system of payment in private health services, some of which charge extra for the additional Kato-Katz procedure. However, the results presented here do not provide enough evidence to advocate that health centres should start using the glass coverslip technique. It seems preferable to deal with the operational aspects leading to the failure of health centres to comply with the present guidelines rather than to introduce a new technique of examination.

Acknowledgements

We thank Mr S. J. De Vlas for his suggestions regarding this manuscript. The Schistosomiasis Control Programme in Burundi is funded by the Burundi Ministry of Health, the Belgian Technical Co-operation, the European Development Fund and the UNDP/World Bank/WHO Special Programme for Research and Training in Tropical Diseases (TDR).

References

Engels, D., Ndoricimpa, J. & Gryseels, B. (1993). Schistosomi-

- asis mansoni in Burundi: progress in its control since 1985. *Bulletin of the World Health Organization*, **71**, 207-214.
- Kato, K. & Miura, M. (1954). Comparative examinations. *Japanese Journal of Parasitology*, **3**, 35.
- Katz, N., Chaves, A. & Pellegrino, J. (1972). A simple device for quantitative stool thick-smear technique in schistosomiasis mansoni. *Revista do Instituto de Medicina Tropical de São Paulo*, **14**, 397-400.
- Peters, P., El Alamy, M., Warren, K. & Mahmoud, A. (1980). Quick Kato smear for field quantification of *Schistosoma mansoni* eggs. *American Journal of Tropical Medicine and Hygiene*, **29**, 217-219.
- Teesdale, C. H. & Amin, M. A. (1976a). A simple thick-smear technique for the diagnosis of *Schistosoma mansoni* infection. *Bulletin of the World Health Organization*, **54**, 703-705.
- Teesdale, C. H. & Amin, M. A. (1976b). Comparison of Bell technique, a modified Kato thick smear technique, and a digestion method for the field diagnosis of schistosomiasis mansoni. *Journal of Helminthology*, **50**, 17-20.
- Teesdale, C. H., Fahringer, K. & Chitsulo, L. (1985). Egg count variability and sensitivity of a thin smear technique for the diagnosis of *Schistosoma mansoni*. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, **79**, 369-373.

Received 25 March 1996; revised 26 April 1996; accepted for publication 30 April 1996

Advertisement

new book

Communicable Disease Epidemiology and Control

Roger Webber,
Department of Epidemiology
and Population Sciences,
London School of Hygiene and
Tropical Medicine,
United Kingdom

Communicable diseases can devastate whole populations and are a problem in both developing countries and the developed world. Understanding their epidemiology is vital to the doctor and communicable disease specialist involved in their control. This book draws on the depth of practical experience gained by the author and a wide range of other sources to review communicable diseases in a global perspective.

The book covers all the important communicable diseases and this is further supported by a comprehensive outline of known communicable diseases given in an annex at the end of the book.

The first part of the book describes epidemiological methods and illustrates their use with practical examples. The second part covers communicable diseases in a systematic manner grouping diseases by epidemiological criteria. This classification enables control to be instigated using the epidemiological principles and control methods described in the first part of the book. Grouping diseases in this manner also makes it easier to understand and link them together, so facilitating learning.

The book balances informativeness with a simple and practical delivery. It is an essential tool for all doctors, epidemiologists, and those working in the control of communicable diseases, especially in developing countries. It is designed to present a clear introduction for medical students, public health specialists and those involved in disease control.

Part I: Theory and methods

- Agent, transmission, host and environment
- Communicable disease theory
- Control principles and strategy
- Control organization
- Control methods
- Notification and health regulations

Part II: Communicable diseases

- Waterwashed diseases
- Faecal-oral diseases
- Soil mediated infections
- Diseases of water contact
- Food borne diseases
- Infectious skin rashes
- Respiratory infections
- Leprosy
- Diseases transmitted via body fluids
- Insect borne diseases
- Ectoparasite zoonoses
- Domestic zoonoses
- Annexes

To place your order for *Communicable Disease Epidemiology and Control*, contact:

Claire Gilman, CAB INTERNATIONAL,
Wallingford, Oxon OX10 8DE, UK
Tel +44 (0) 1491 832111 Fax +44 (0) 1491 826090
e-mail c.gilman@cabi.org

July 1996 368 pages (Paperback) ISBN 0 85199 138 6 Price: £19.95 (US\$37.50 Americas only)